



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of)	
Jean-Marc Balloul et al.)	Group Art Unit: 1648
Application No.: 09/506,942)	Examiner: Shanon A. Foley
Filed: February 18, 2000)	Confirmation No.: 9626
For: PHARMACEUTICAL)	
COMPOSITION FOR TREATING)	
PAPILLOMAVIRUS TUMORS AND)	
INFECTION)	

DECLARATION BY JEAN-MARC BALLOUL UNDER 37 C.F.R. § 1.131

Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

Sir:

I, Jean-Marc Balloul, hereby state as follows:

1. I am a named inventor on U.S. Application No. 09/506,942 ("the '942 application").
2. I believe and allege the '942 application is a divisional application of U.S. Application Serial No. 09/043,933 ("the '933 application"), filed March 30, 1998, which claims priority benefit of French Application No. 96-09584, filed July 30, 1996.
3. I believe and allege we conceived the invention claimed in the '942 application prior to March 1996, and that conception was pursued with diligence from the time it was conceived before March 1996 until we filed the priority application, French Application No. 96-09584, on July 30, 1996.
4. Prior to March 1996, I participated in preparing an internal report for a scientific council at Transgene, attached as Exhibit A. This report discusses the production of clinical batches of vaccina virus construct VVTG5021 & 5065. This report further demonstrates expression of HPV genes and IL-2, the absence of

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toxicity seen in animal models following injection. The report also provides the same immunoprotein and immunotherapy assays as seen in Example 6 of the '942 application. The exact date of the internal report has been redacted on the copy provided. Further, data not connected to the '942 application has also been redacted on the copy provided.

5. Exhibit A, and the work detailed therein, was prepared in France, a World Trade Organization (WTO) member country, prior to March 1996.

6. We believe and allege each element of the claims of the '942 application is disclosed in Exhibit A.

7. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Date:

April 6th, 2006



JEAN-MARC BALLOUL

EXHIBIT A

Scientific Council. TRANSGENE SA

Project N°87

Research and Development

Scientific Direction: MPK

Project Leader: JMB

Molecular Biology and Virology: MG, KD, ReB, JMB

Pre-clinical studies: NaB, CHP, LS, GiG, LR, BA

Production and controls: KD, EB, OF, EK, DV, HeL, JMB

Cervical cancer associated with Human Papillomavirus

Among the 75 types of human papillomavirus types so far identified 20 distinct isolates are known to infect the genital tract and 4 types (HPV16, 18, 31 and 45) account for 80% of HPV infection associated with cervical cancer. Although cervical cancer rates have been declining, they remain relatively high in developing country where prevention is lacking. Both therapeutic vaccination (induction of the regression of precancerous and cancerous lesions associated with oncogenic HPV types), and prophylactic vaccination are under investigation in various laboratories throughout the world. Prophylaxis with HPV vaccines is certainly the most promising approach and will have a long term impact in reducing cervical cancer. At the moment a consensus exist for the choice of the immunogens to be include in prophylactic vaccines. They are in fact the virus-like particles (VLP) synthesised *in vitro*. These DNA free particles shown their ability to induce neutralizing antibodies. Several compaignies are now proceeding to the development and production of VLPs at the standards required for human trials.

Clinical batch for HPV16 invasive carcinoma immunotherapy

As we predict that successful vaccination against the human papillomavirus requires induction of both cytotoxic T lymphocyte (CTL) response against early viral proteins and neutralizing antibodies against capsid proteins L1/L2 to prevent reinfection we developed a vaccinia virus expressing both early and late HPV antigens under the control of early late vaccinia virus promoters. Briefly HPV16 E7 and E6 have been mutated according the litterature in order to abrogate their transforming activity. Then mutant genes were inserted by homologuous recombination at the K1L locus of the vaccinia virus genome (Copenhagen strain) and placed under the control of the early late vaccinia virus promoter H5R. At the same locus we inserted both the human Il2 gene placed under the control of the early late vaccinia virus promoter p7.5 and a selection marker gene LacZ placed under the control of the K1L promoter (see figure 1). Genes encoding for the late viral proteins L1 and L2 were inserted into the TK locus of the vaccinia virus genome and placed under the control of the vaccinia virus early-late promoter p7.5 (see figure 2).

Clinical batch was produced in our L3 unit on primary CEF. The viral amplification leads to the production of more than 3,000 doses of vaccines containing around 10E8 pfu per doses. Expression of HPV genes was analyzed by Western blot analysis. As shown in figure 3, 4,5 6, HPV antigens are expressed and detectable by Western blot. Level of Il2 was measured and evaluated to 10ng/ml/24h/10E6 cells infected with 1pfu per cell (see figure 7). Complete sequencing of open reading frames showed in the case of Il2 the accidental cloning of a 600bp

DNA fragment corresponding to the β -globin intron present in the parental plasmid. As 4 ORFs encoding for small polypeptides (26 to 56 aa) could be taken in account by the early-late vaccinia virus promoter controlling the expression of the human IL2 gene we analyzed the presence of such contaminants in VVTG5021&5065 infected BHK-21 extracts. Results showed that no contaminants are detectable compared with VVTG5021&5061 extracts expressing all genes present in the VVTG5021&5065 except the human IL2 gene (see figures 8, 9).

New formulation of HPV16 early antigens in vaccinia virus

MUC1/B7.1 formulation

In order to enhanced immunogenicity of the MUC-1 antigen we constructed a new replicative vaccinia virus co-expressing the human B7.1 gene and the human MUC-1 cDNA (name code pTG6005 figure 13). MUC-1 cDNA available at Transgene contains more than 24 tandem repeats and regarding the propensity of the vaccinia virus genome to eliminate DNA repetitions we analyzed number of plaques after the first TK- selection. We isolate one clone containing at least 12 repeat units which is an impressive number compared to the VVTG5058 viral isolate.

Non Replicative Vaccinia Virus as Vector for Cancer Immunotherapy

Introduction

Vaccinia virus is the prototype of Orthopoxvirus. It can replicate in a wide range of cells. At least three viral genes affect the host range of orthopoxviruses in tissue culture: K1L gene, C7L gene and the cowpox 77-kDa gene. As regulatory authorities showed some fair concerning the use of replicative vectors for human health, we decided to develop attenuated poxvirus expression vectors. First question was to select the appropriate strain of vaccinia virus. Paolletti et al. has previously and extensively described various pox virus in their non replicative form such as NYCAC (highly attenuated vaccinia virus engineered from the Copenhagen strain) and ALVAC (Avian canaripox which is not replicative into mamalian cells). Unfortunately vectors for homologous recombination into these non replicative poxvirus were not accessible for Transgene. In the same time G. Sutter in B. Moss laboratory (Laboratory of viral diseases, National Institute of Allergy and Infectious Diseases, NIH, Bethesda, USA) has described a recombinant vector derived from the host range-restricted and highly attenuated MVA strain of vaccinia virus which has been shown to express the inserted foreign gene in authentic fashion and to provide the antigen to the immune system in such manner that it induces a protective immunity against pathogen such like the influenza virus in mice when it expresses haemagglutinin and nucleoprotein genes.

MVA known as modified vaccinia virus Ankara was found to be avirulent in normal or immunosuppresses animals ranging from rodents to macaques and was without significant side-effects in 120,000 humans, many of whom were at high risk for conventional smallpox vaccine. During over 570 passages in chicken embryo fibroblasts, MVA became host-restricted and unable to grow in almost all tested mamalian cell lines including those of rodent and human origin. Analysis of the genome revealed that viral DNA had suffered six major deletions resulting in the loss of 30000 base pairs equivalent of 15% of its genetic information. The replication of MVA DNA implied that the initial stages of infection comprising viral attachment, entry, early gene expression, and uncoating occurred in non permissive human cells. G. Sutter et al. has observed that only immature virus particles appearing as circular spiculecoated membranes encircling granular material and lacking dense nucleoprotein bodies could be seen by electron microscopic examination of human cells infected with MVA whereas mature brick-shaped particles with complex internal structures were numerous in human cells with WT or CEF with MVA. Moreover no increase in MVA titer was detected in Hela or 293 cell lines, and

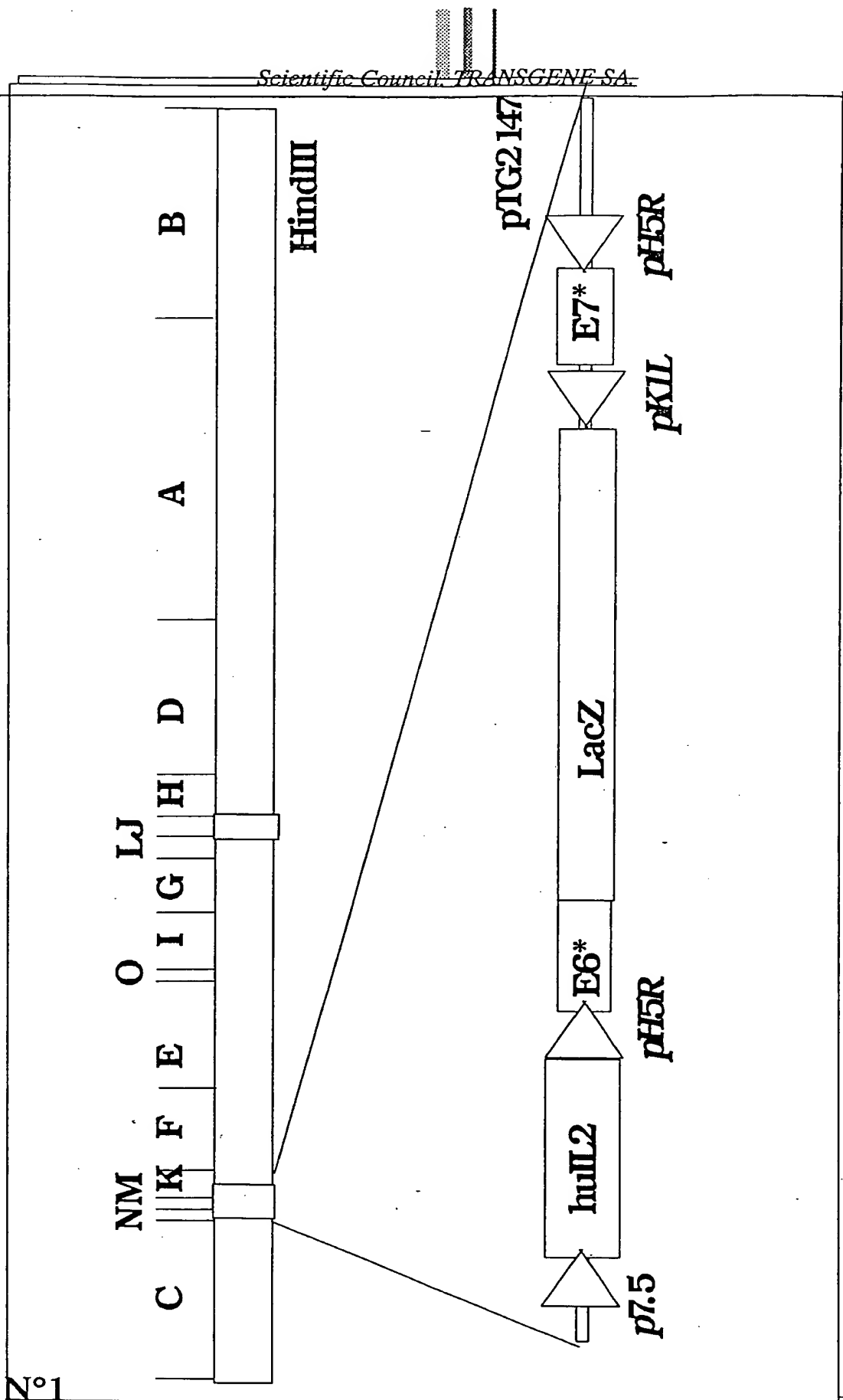
at the opposite WT increase 10,000-fold in titer in the same conditions. These differences are not associated with DNA replication because concatemeric forms of replicative MVA DNA are processes normally to unit genome. These authors observed common viral protein pattern for both WT and MVA in infected human cells with one exception for a 90kDa polypeptide which is not seen in the MVA protein pattern. This is attributed to the deletion of the gene encoding nonessential A-type inclusion protein homolog. In pulse-chase assays they also showed that if in WT infected Hela cells expected cleavage of at least 5 polypeptides including the major core protein precursors P4a and P4b occurred, the processing of MVA proteins was inhibited.

Plasmid construction

PCR segments flanking either deletion III or deletion II were cloned into pTG1E at the unique EcoRI site (code name pTG6018 containing flanking DNA segments for homologous recombination at the deletion II site and pTG6019 containing flanking DNA segments for homologous recombination at the deletion III site figure 14). In order to facilitate the isolation of recombinant viruses we inserted between these flanks a double expression cassette composed of the early late promoter p7.5 controlling the marker gene and of the early late promoter p7.5 in the same orientation controlling a reporter gene for internal recombination which encode for the human Il2. We have selected as markers either the LacZ gene encoding β -galactosidase (β -gal) in the case of deletion II and the E. coli *gusA* gene encoding β -glucuronidase (GUS) in the case of the deletion III. β -gal and GUS being from E. coli origin we decided to eliminate these genes from the final clinical product. For these purpose we used the natural propensity of the vaccinia virus to eliminate DNA repeated sequences. Therefore p7.5 flanking marker gene should recombine properly, excising the E. Coli gene and reconstituting a functional p7.5 promoter controlling the human Il-2 gene (figure 15).

VVTG5065

Figure N°1



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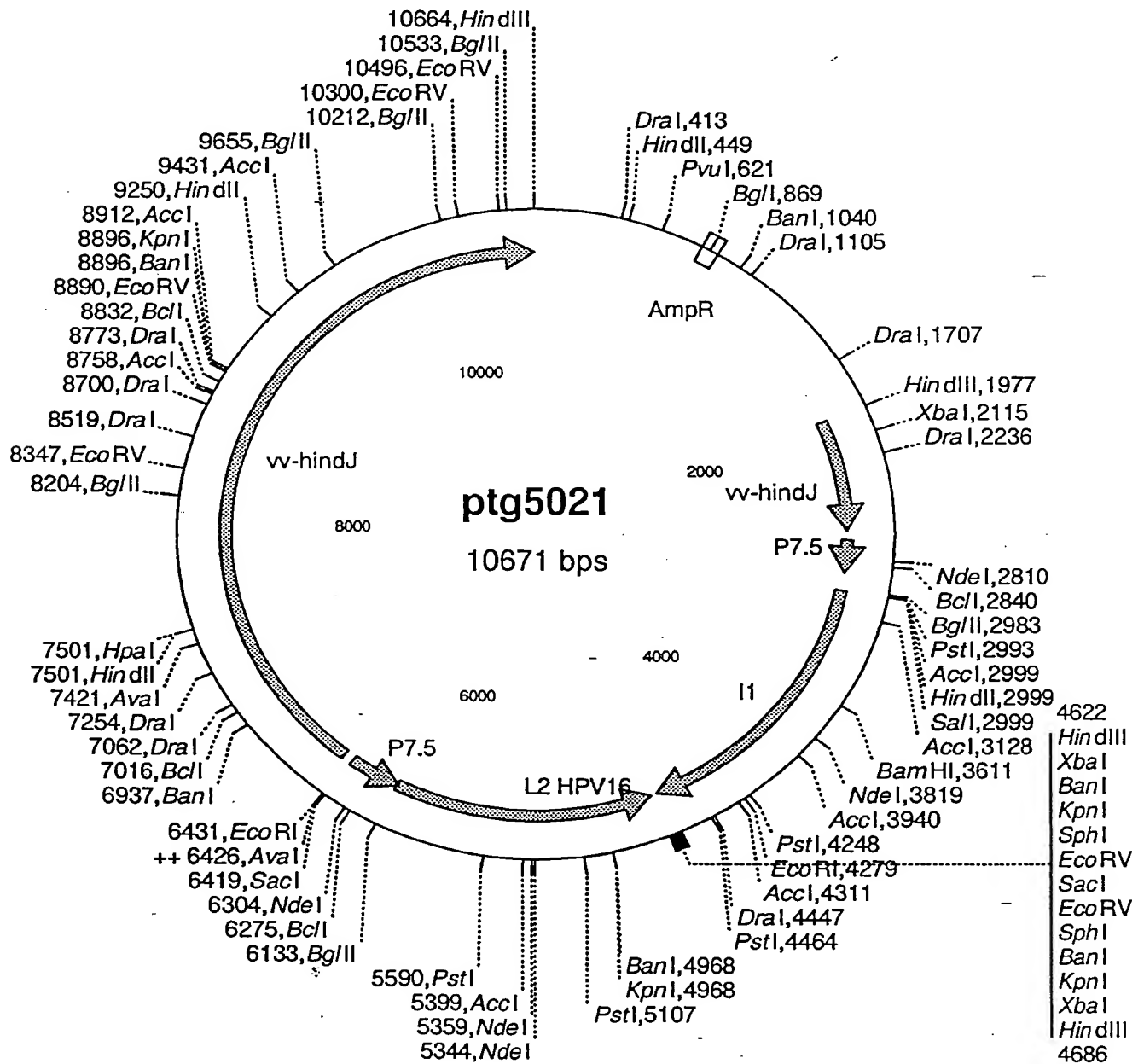
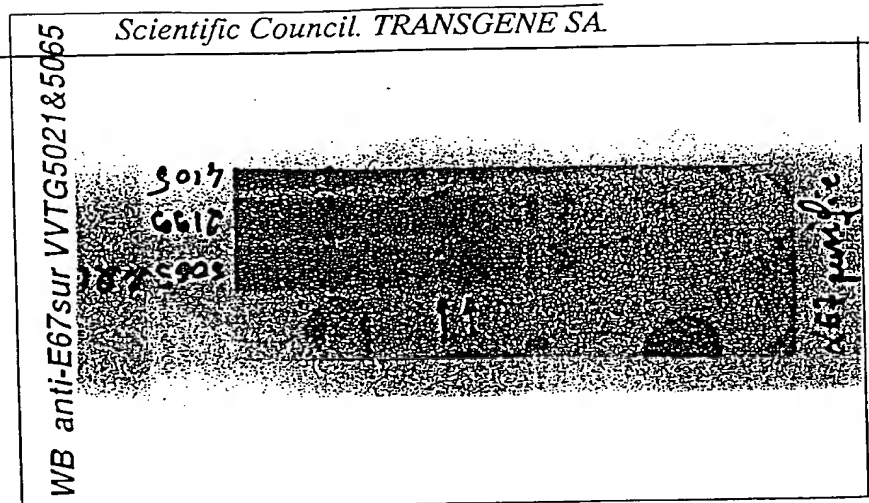


Figure N°2

Figure N°4



Séquence pTG5021&5065

DNA Align (ALIGN) version 3.90 Apr 93
this is Align ~5085 SEQ(1,6829) with E7-SEQ(1,297) ktuple: 3 Gap penalty: 3 Largest: 20

Alignment of: 6829 bases from 1 to 6829

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VERSUS
2007.06.01 to 2007.06.01

Gap penalty = 3

range = 20 gap penalty = 3
k-tuple size = 3
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110V 120V 130V 140V 150V 160V 170V 180V 190V 200V
ATGCTGAGGACAC
ATGCTGAGGACAC

10°	280v	780v	10°
270v	460v	270v	460v
250v	350v	250v	350v
230v	310v	230v	310v
210v	270v	210v	270v
190v	230v	190v	230v
170v	190v	170v	190v
150v	150v	150v	150v
130v	110v	130v	110v
110v	70v	110v	70v
90v	30v	90v	30v
70v	0v	70v	0v
50v		50v	
30v		30v	
10v		10v	
0v		0v	
10°	280v	780v	10°
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210v	270v	210v	270v
190v	230v	190v	230v
170v	190v	170v	190v
150v	150v	150v	150v
130v	110v	130v	110v
110v	70v	110v	70v
90v	30v	90v	30v
70v	0v	70v	0v
50v		50v	
30v		30v	
10v		10v	
0v		0v	
10°	280v	780v	10°
270v	460v	270v	460v
250v	350v	250v	350v
230v	310v	230v	310v
210v	270v	210v	270v
190v	230v	190v	230v
170v	190v	170v	190v
150v	150v	150v	150v
130v	110v	130v	110v
110v	70v	110v	70v
90v	30v	90v	30v
70v	0v	70v	0v
50v		50v	
30v		30v	
10v		10v	
0v		0v	

$\log_{10} \text{base ant } 1000000$ & $\log_{10} = 279$

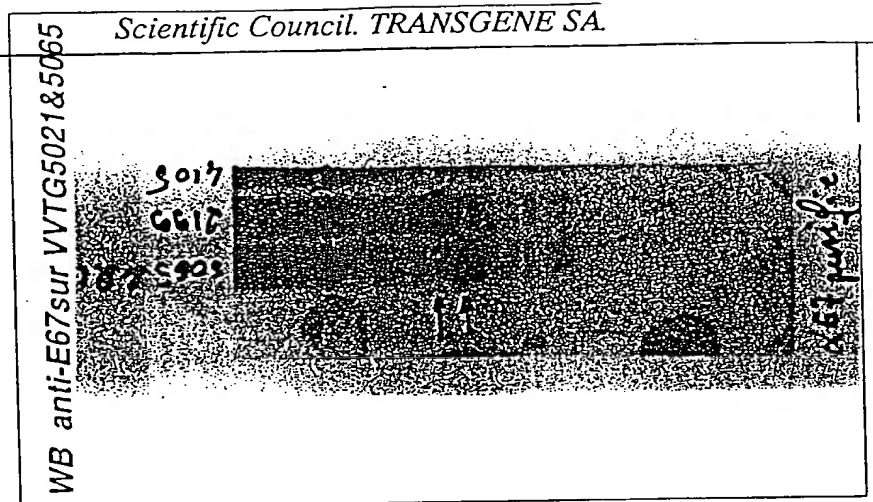
Similarity index = 99.643%, total bases not developed = 1, bases non-site gaps = 18

Number of gaps = 1, bases opposite gaps = 29

Length of the overlapping region = 291
Number of unambiguous matching bases = 279

Number of unambiguous matching bases = 0
Number of ambiguous matching bases = 0

Figure N°4



Séquence pTG5021&5065

DNA Align (ALIGN) version 3.90 Apr 93
 ...SOL5 SEQID AB291 with E7-SEQ(1,297) ktuple= 3 Gap penalty: 3 Largest: 20

Alignment of: 6829 bases from 1 to 6829

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C:\JMB\CANUTER\PL5065\1E7.SEO 297 bases from 1 (0.4%)

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 ATGCGATGAGATACACC

57

[illegible]

~5065
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CAATTAAATGACAGCTCAGAGGAGGAGGATGAAATAGAT

TACATTGTCATGAATATATGTGGTATTTGCAGCCAGAGACAACCTGATCTCTACTGTTATGACCAATTAAATGACAGTCL	70°	80°	90°	100°	110°
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380V 370V 360V 350V 340V 330V 320V 310V 300V

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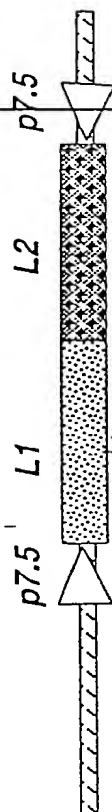
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length of the overlapping region = 297

Length of the unambiguous matching bases = 279
Number of unambiguous matching bases = 0

Number of ambiguous matching bases = 0

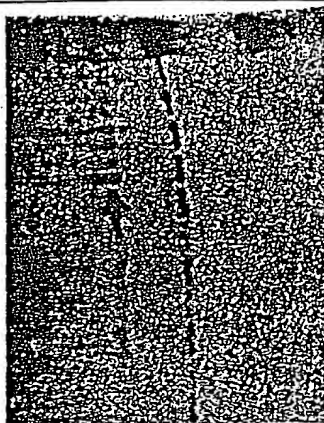
Figure N°5

L1 produite par
VVTG5021&5065

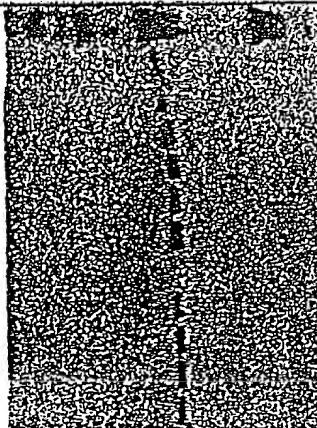


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WB/ICAM-1 sur VVTG5021&5065



WTG5021
59056



14

△ A Sacros 2

Eval10.95/87/1311

CIGIACGATTCGTATGATGGTAGTGCTTTTCTGCTTGCGATTATTAAGTCATCAGTGTGCTG

V 7780W 7790W 7800W 7810W 7820W 7830W 7840W 7850W

Figure N°6

-PPL16	4230v	4240v	4250v	4260v	4270v	4280v	4290v	4300v	4310v
L2-5021	4320v	4330v	4340v	4350v	4360v	4370v	4380v	4390v	4400v
-PPL16	4410v	4420v	4430v	4440v	4450v	4460v	4470v	4480v	4490v
L2-5021	4500v	4510v	4520v	4530v	4540v	4550v	4560v	4570v	4580v
-PPL16	4590v	4600v	4610v	4620v	4630v	4640v	4650v	4660v	4670v
L2-5021	4680v	4690v	4700v	4710v	4720v	4730v	4740v	4750v	4760v
-PPL16	4770v	4780v	4790v	4800v	4810v	4820v	4830v	4840v	4850v
L2-5021	4860v	4870v	4880v	4890v	4900v	4910v	4920v	4930v	4940v
-PPL16	4950v	4960v	4970v	4980v	4990v	5000v	5010v	5020v	5030v
L2-5021	5040v	5050v	5060v	5070v	5080v	5090v	5100v	5110v	5120v
-PPL16	5130v	5140v	5150v	5160v	5170v	5180v	5190v	5200v	5210v
L2-5021	5220v	5230v	5240v	5250v	5260v	5270v	5280v	5290v	5300v
-PPL16	5310v	5320v	5330v	5340v	5350v	5360v	5370v	5380v	5390v
L2-5021	5400v	5410v	5420v	5430v	5440v	5450v	5460v	5470v	5480v
-PPL16	5490v	5500v	5510v	5520v	5530v	5540v	5550v	5560v	5570v
L2-5021	5580v	5590v	5600v	5610v	5620v	5630v	5640v	5650v	5660v
-PPL16	5670v	5680v	5690v	5700v	5710v	5720v	5730v	5740v	5750v
L2-5021	5760v	5770v	5780v	5790v	5800v	5810v	5820v	5830v	5840v
-PPL16	5850v	5860v	5870v	5880v	5890v	5900v	5910v	5920v	5930v
L2-5021	5940v	5950v	5960v	5970v	5980v	5990v	6000v	6010v	6020v
-PPL16	6030v	6040v	6050v	6060v	6070v	6080v	6090v	6100v	6110v
L2-5021	6120v	6130v	6140v	6150v	6160v	6170v	6180v	6190v	6200v
-PPL16	6210v	6220v	6230v	6240v	6250v	6260v	6270v	6280v	6290v
L2-5021	6300v	6310v	6320v	6330v	6340v	6350v	6360v	6370v	6380v
-PPL16	6390v	6400v	6410v	6420v	6430v	6440v	6450v	6460v	6470v
L2-5021	6480v	6490v	6500v	6510v	6520v	6530v	6540v	6550v	6560v
-PPL16	6570v	6580v	6590v	6600v	6610v	6620v	6630v	6640v	6650v
L2-5021	6660v	6670v	6680v	6690v	6700v	6710v	6720v	6730v	6740v
-PPL16	6750v	6760v	6770v	6780v	6790v	6800v	6810v	6820v	6830v
L2-5021	6840v	6850v	6860v	6870v	6880v	6890v	6900v	6910v	6920v
-PPL16	6930v	6940v	6950v	6960v	6970v	6980v	6990v	7000v	7010v
L2-5021	7020v	7030v	7040v	7050v	7060v	7070v	7080v	7090v	7100v
-PPL16	7110v	7120v	7130v	7140v	7150v	7160v	7170v	7180v	7190v
L2-5021	7200v	7210v	7220v	7230v	7240v	7250v	7260v	7270v	7280v
-PPL16	7290v	7300v	7310v	7320v	7330v	7340v	7350v	7360v	7370v
L2-5021	7380v	7390v	7400v	7410v	7420v	7430v	7440v	7450v	7460v
-PPL16	7470v	7480v	7490v	7500v	7510v	7520v	7530v	7540v	7550v
L2-5021	7560v	7570v	7580v	7590v	7600v	7610v	7620v	7630v	7640v
-PPL16	7650v	7660v	7670v	7680v	7690v	7700v	7710v	7720v	7730v
L2-5021	7740v	7750v	7760v	7770v	7780v	7790v	7800v	7810v	7820v
-PPL16	7830v	7840v	7850v	7860v	7870v	7880v	7890v	7900v	7910v
L2-5021	7920v	7930v	7940v	7950v	7960v	7970v	7980v	7990v	8000v
-PPL16	8010v	8020v	8030v	8040v	8050v	8060v	8070v	8080v	8090v
L2-5021	8100v	8110v	8120v	8130v	8140v	8150v	8160v	8170v	8180v
-PPL16	8190v	8200v	8210v	8220v	8230v	8240v	8250v	8260v	8270v
L2-5021	8280v	8290v	8300v	8310v	8320v	8330v	8340v	8350v	8360v
-PPL16	8370v	8380v	8390v	8400v	8410v	8420v	8430v	8440v	8450v
L2-5021	8460v	8470v	8480v	8490v	8500v	8510v	8520v	8530v	8540v
-PPL16	8550v	8560v	8570v	8580v	8590v	8600v	8610v	8620v	8630v
L2-5021	8640v	8650v	8660v	8670v	8680v	8690v	8700v	8710v	8720v
-PPL16	8730v	8740v	8750v	8760v	8770v	8780v	8790v	8800v	8810v
L2-5021	8820v	8830v	8840v	8850v	8860v	8870v	8880v	8890v	8900v
-PPL16	8910v	8920v	8930v	8940v	8950v	8960v	8970v	8980v	8990v
L2-5021	9000v	9010v	9020v	9030v	9040v	9050v	9060v	9070v	9080v
-PPL16	9090v	9100v	9110v	9120v	9130v	9140v	9150v	9160v	9170v
L2-5021	9180v	9190v	9200v	9210v	9220v	9230v	9240v	9250v	9260v
-PPL16	9270v	9280v	9290v	9300v	9310v	9320v	9330v	9340v	9350v
L2-5021	9360v	9370v	9380v	9390v	9400v	9410v	9420v	9430v	9440v
-PPL16	9450v	9460v	9470v	9480v	9490v	9500v	9510v	9520v	9530v
L2-5021	9540v	9550v	9560v	9570v	9580v	9590v	9600v	9610v	9620v
-PPL16	9630v	9640v	9650v	9660v	9670v	9680v	9690v	9700v	9710v
L2-5021	9720v	9730v	9740v	9750v	9760v	9770v	9780v	9790v	9800v
-PPL16	9810v	9820v	9830v	9840v	9850v	9860v	9870v	9880v	9890v
L2-5021	9900v	9910v	9920v	9930v	9940v	9950v	9960v	9970v	9980v
-PPL16	9990v	10000v	10010v	10020v	10030v	10040v	10050v	10060v	10070v
L2-5021	10080v	10090v	10100v	10110v	10120v	10130v	10140v	10150v	10160v
-PPL16	10170v	10180v	10190v	10200v	10210v	10220v	10230v	10240v	10250v
L2-5021	10260v	10270v	10280v	10290v	10300v	10310v	10320v	10330v	10340v
-PPL16	10350v	10360v	10370v	10380v	10390v	10400v	10410v	10420v	10430v
L2-5021	10440v	10450v	10460v	10470v	10480v	10490v	10500v	10510v	10520v
-PPL16	10530v	10540v	10550v	10560v	10570v	10580v	10590v	10600v	10610v
L2-5021	10620v	10630v	10640v	10650v	10660v	10670v	10680v	10690v	10700v
-PPL16	10710v	10720v	10730v	10740v	10750v	10760v	10770v	10780v	10790v
L2-5021	10800v	10810v	10820v	10830v	10840v	10850v	10860v	10870v	10880v
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L2-5021	10980v	10990v	11000v	11010v	11020v	11030v	11040v	11050v	11060v
-PPL16	11070v	11080v	11090v	11100v	11110v	11120v	11130v	11140v	11150v
L2-5021	11160v	11170v	11180v	11190v	11200v	11210v	11220v	11230v	11240v
-PPL16	11250v	11260v	11270v	11280v	11290v	11300v	11310v	11320v	11330v
L2-5021	11340v	11350v	11360v	11370v	11380v	11390v	11400v	11410v	11420v
-PPL16	11430v	11440v	11450v	11460v	11470v	11480v	11490v	11500v	11510v
L2-5021	11520v	11530v	11540v	11550v	11560v	11570v	11580v	11590v	11600v
-PPL16	11610v	11620v	11630v	11640v	11650v	11660v	11670v	11680v	11690v
L2-5021	11700v	11710v	11720v	11730v	11740v	11750v	11760v	11770v	11780v
-PPL16	11790v	11800v	11810v	11820v	11830v	11840v	11850v	11860v	11870v
L2-5021	11880v	11890v	11900v	11910v	11920v	11930v	11940v	11950v	11960v
-PPL16	11970v	11980v	11990v	12000v	12010v	12020v	12030v	12040v	12050v
L2-5021	12060v	12070v	12080v	12090v	12100v	12110v	12120v	12130v	12140v
-PPL16	12150v	12160v	12170v	12180v	12190v	12200v	12210v	12220v	12230v
L2-5021	12240v	12250v	12260v	12270v	12280v	12290v	12300v	12310v	12320v
-PPL16	12330v	12340v	12350v	12360v	12370v	12380v	12390v	12400v	12410v
L2-5021	12420v	12430v	12440v	12450v	12460v	12470v	12480v	12490v	12500v
-PPL16	12510v	12520v	12530v	12540v	12550v	12560v	12570v	12580v	12590v
L2-5021	12600v	12610v	12620v	12630v	12640v	12650v	12660v	12670v	12680v
-PPL16	12690v	12700v	12710v	12720v	12730v	12740v	12750v	12760v	12770v
L2-5021	12780v	12790v	12800v	12810v	12820v	12830v	12840v	12850v	12860v
-PPL16	12870v	12880v	12890v	12900v	12910v	12920v	12930v	12940v	12950v
L2-5021	12960v	12970v	12980v	12990v	13000v	13010v	13020v	13030v	13040v
-PPL16	13050v	13060v	13070v	13080v	13090v	13100v	13110v	13120v	13130v
L2-5021	13140v	13150v	13160v	13170v	13180v	13190v	13200v	13210v	13220v
-PPL16	13230v	13240v	13250v	13260v	13270v	13280v	13290v	13300v	13310v
L2-5021	13320v	13330v	13340v	13350v	13360v	13370v	13380v	13390v	13400v
-PPL16	13410v	13420v	13430v	13440v	13450v	13460v	13470v	13480v	13490v
L2-5021	13500v	13510v	13520v	13530v	13540v	13550v	13560v	13570v	13580v
-PPL16	13590v	13600v	13610v	13620v	13630v	13640v	13650v	13660v	13670v
L2-5021	13680v	13690v	13700v	13710v	13720v	13730v	13740v	13750v	13760v
-PPL16	13770v	13780v	13790v	13800v	13810v	13820v	13830v	13840v	13850v
L2-5021	13860v	13870v	13880v	13890v	13900v	13910v	13920v	13930v	13940v
-PPL16	13950v	13960v	13970v	13980v	13990v	14000v	14010v	14020v	14030v
L2-5021	14040v	14050v	14060v	14070v	14080v	14090v	14100v	14110v	14120v
-PPL16	14130v	14140v	14150v	14160v	14170v	14180v	14190v	14200v	14210v
L2-5021	14220v	14230v	14240v	14250v	14260v	14270			

Figure N°7

I12 produite par VVTG5021&5065

L2 p7.5

RESULTATS MANIP ELISA IL2E9506 du 25.01.95

	standard R&D	NIBSC = R&Dx1.55	standard Boehringer
SN VVTG 5058 lot brut = préstock	421,0 ng/ml	652,6 ng/ml	612,0 ng/ml
SN VVTG 5058 stock	545,2 ng/ml	845,1 ng/ml	769,9 ng/ml
SN VVTG 5058 lot purifié	357,2 ng/ml	553,7 ng/ml	518,1 ng/ml
SN ETAm	0,002 ng/ml	0,003 ng/ml	0,006 ng/ml
SN VVTG 65X21	4,5 ng/ml	7,0 ng/ml	7,7 ng/ml
SN VVTG 61X21	0,003 ng/ml	0,005 ng/ml	0,008 ng/ml
SN WT	0,011 ng/ml	0,017 ng/ml	0,02 ng/ml

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	standard Boehringer
SN VVTG 5058 lot brut = préstock	798,4 ng/ml
SN VVTG 5058 stock	902,8 ng/ml
SN VVTG 5058 lot purifié	807,5 ng/ml
SN ETAm	0 ng/ml
SN VVTG 65X21	5,7 ng/ml
SN VVTG 61X21	0 ng/ml
SN WT	0 ng/ml

Eval10.95/87/184

Séquence pTG5021&5065

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

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Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

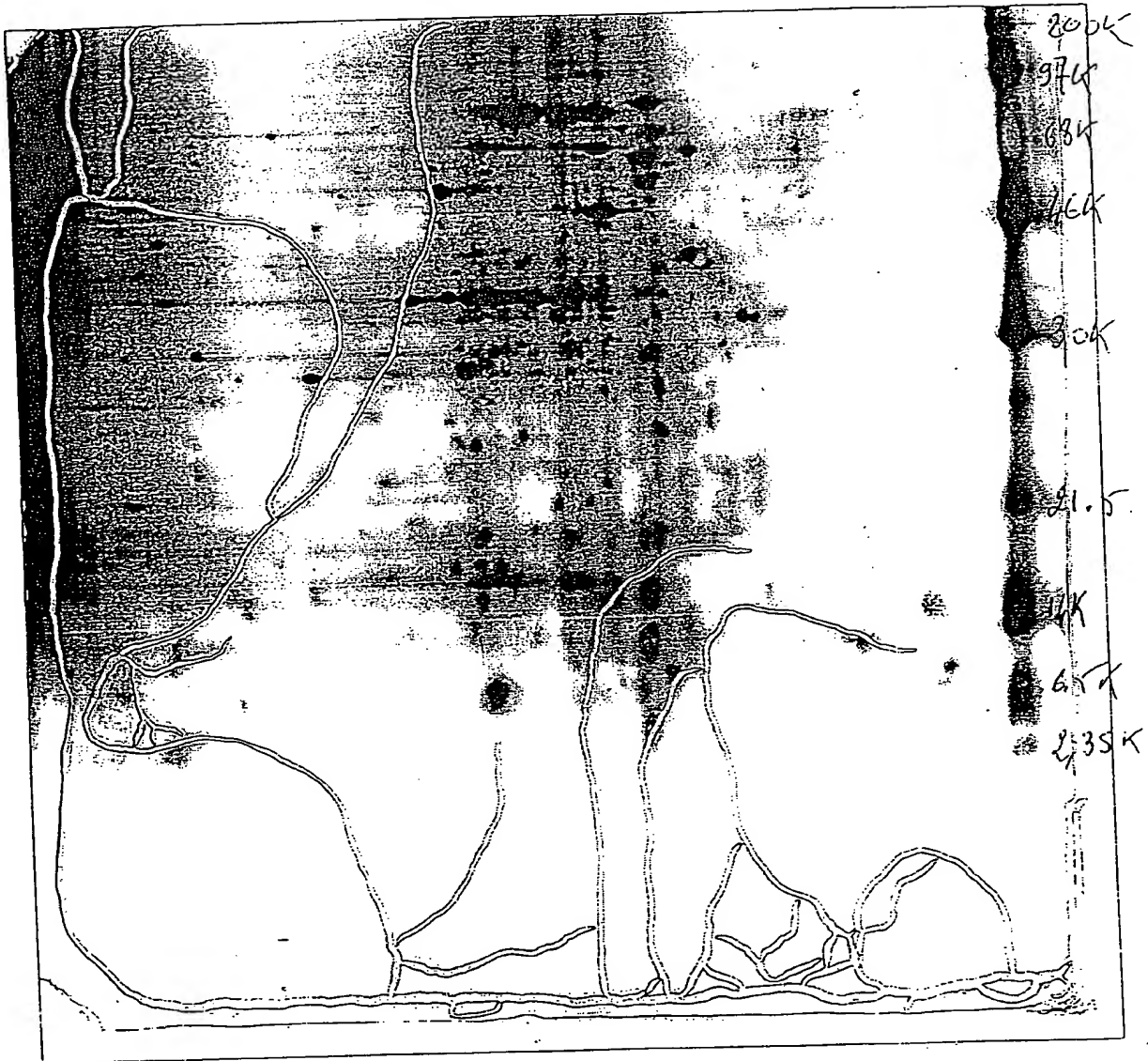
Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

Alignement de 5065 bases de 1 à 2750

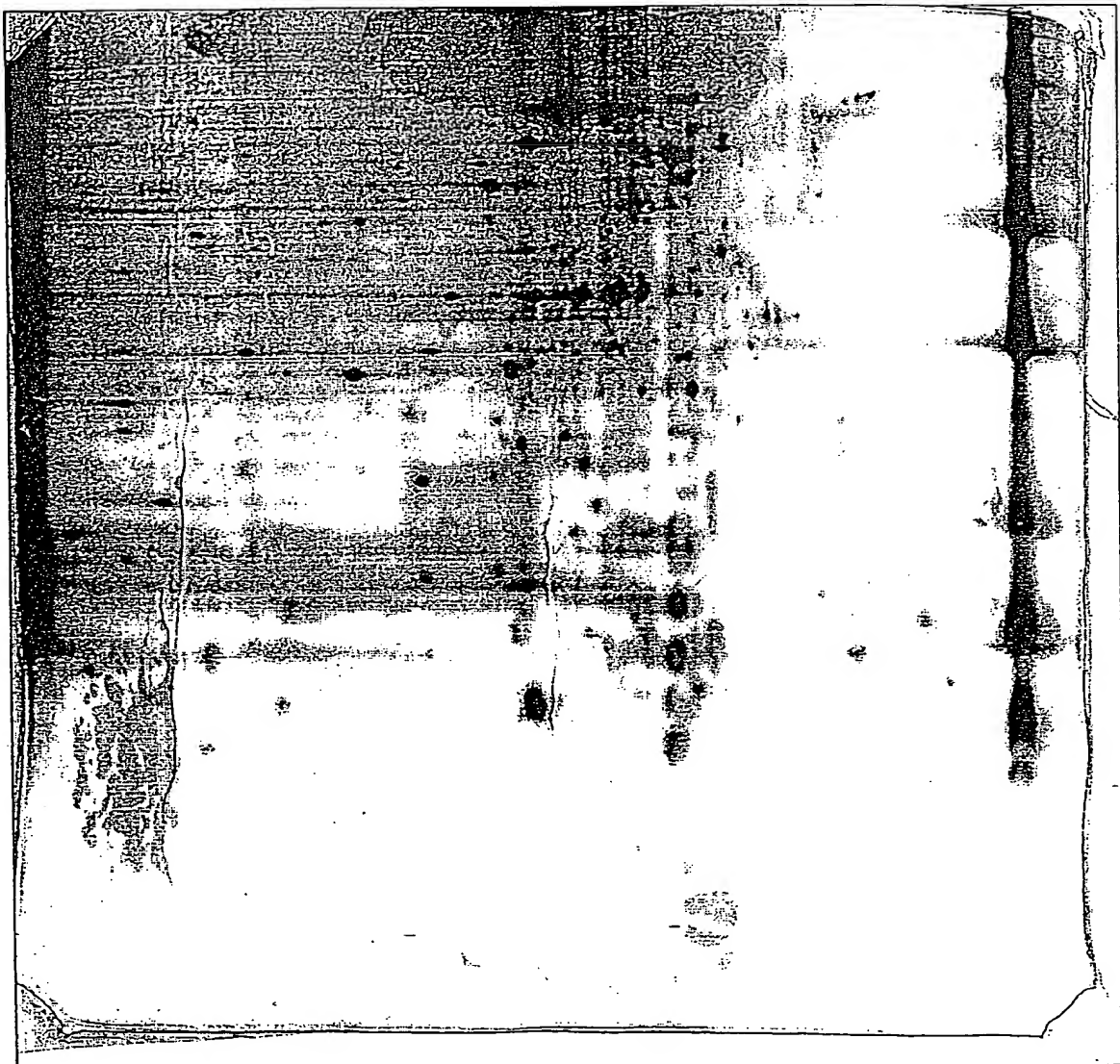
Alignement de 5065 bases de 1 à 2750



Nitrate d'argent, fixation des
petites protéines par la
formaldéhyde

VVTG5021&5061

Scientific Council TRANSGENE SA.



Nitrate d'argent, fixation des
petits peptides par la
formaldéhyde

Figure N°9

Eval.95/87/131/132

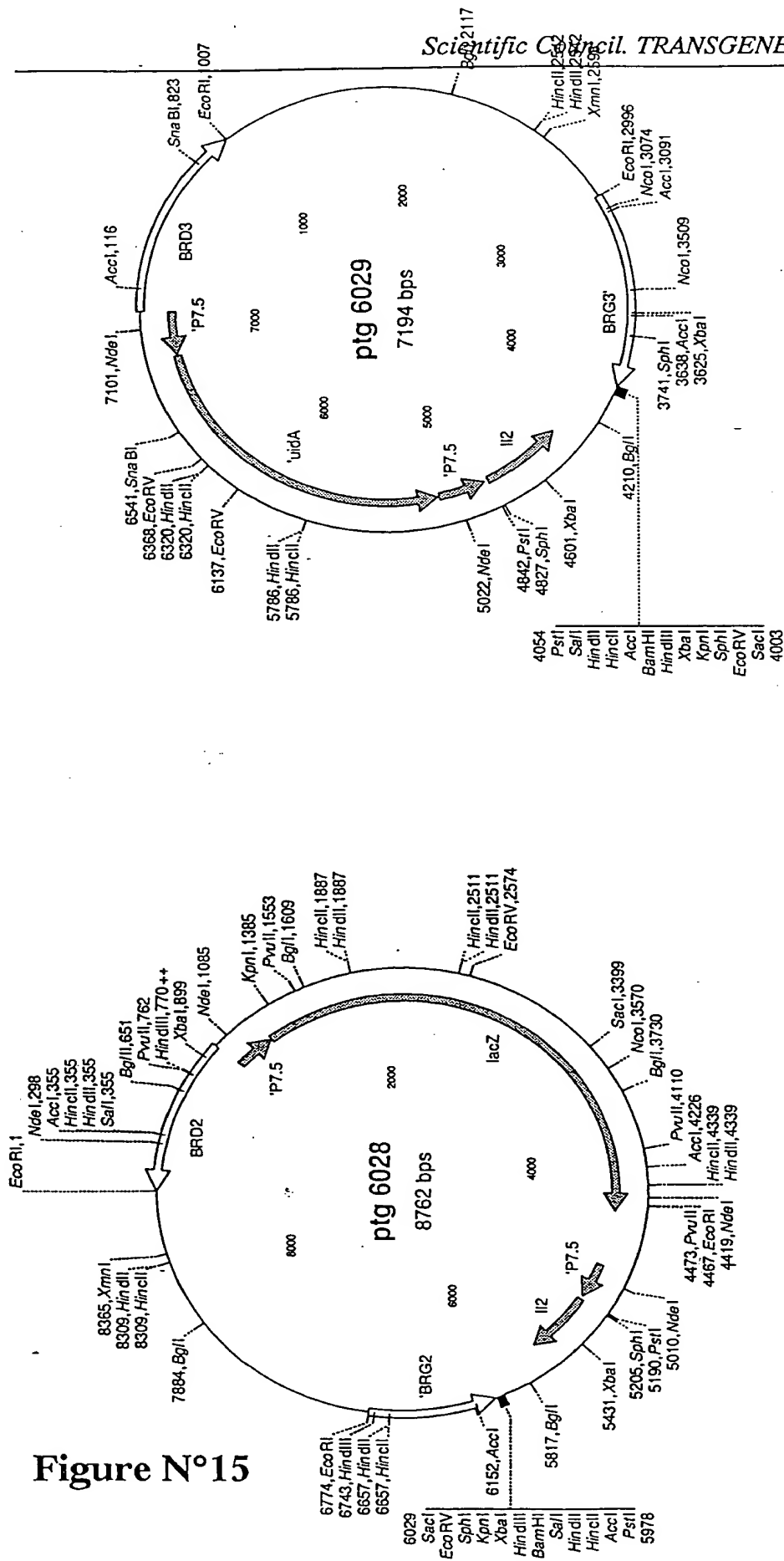


Figure N°15

Nadine Bizouarne, Catherine Pêcheur

Scientific council,

Project 87 : Animal models for vaccinia based vectors for specific cancer immunotherapy.

Infection of mucosal epithelium by papillomaviruses plays a critical role in the development of genital and oral warts and is implicated in the induction of human oropharyngeal and cervical cancer. There are actually no vaccine to prevent disease caused by HPV infection. Different levels of problems are encountered in the development of vaccine strategies. First, these viruses are highly species-specific and their *in vitro* culture is, up to now, impossible, making the use of a "simple" animal model out of the question. All the experiments described in this report were done in an "artificial" animal model consisting of mice grafted with tumor cells expressing an HPV target molecule (E7). Second, the HPV uses several strategies to escape its host immune system. In particular i) mucosal lesions caused by HPV induce small quantities of viral particle (the induction of an humoral immune response seems to be only complementary to a cellular one), ii) during the transformation event of the host cell, the viral DNA is integrated to the cellular genome and only the viral early proteins continue to be expressed at low levels, reducing our choice for antigen-specific immunotherapy. As described previously by JM Balloul, a recombinant vaccinia virus expressing E6*E7*L1L2.L2 (non oncogenic mutated forms of E6 and E7 proteins, VV5021X5065) was produced. In this report we will describe the *in vivo* activities of this vaccinia vector and several approaches used to obtain a second generation vaccinia virus vector.

1- *In vivo* toxicity of VV5021X5065 :

In order to demonstrate the attenuation of the VV5021X5065, we compared the toxicity of this construct to the toxicity of a wild type vaccinia virus. This comparison was made in immunodeficient mice.

1.1 Dose effect : Nude mice were injected intramuscularly with 10^6 , 10^7 or 10^8 pfu of VV5021X5065 or VVwt. The number of mice developing vaccinia lesions are indicated in the table 1, and show that even when injected with the non attenuated VVwt, few mice have vaccinia lesions.

1.2 Immunization route effect : Nude mice were injected with 10^7 pfu of vaccinia by different routes of immunization : intracranial , intramuscular, intravenous, or subcutaneous. The table 2 represents the number of mice having vaccinia lesions:

It seems clear that the VV5021X5065 is consistently attenuated comparing to the VVwt: even when injected by the intracranial route, this recombinant does not lead to vaccinia lesions in this experimental system.

VV	WT	IV	5/5	VV5021X5065	0/5
		IC	4/5		0/5
		IP	3/5		0/5
		TH	1/5		0/5

... VV recombinant attenuated

2- *In vivo* antiviral effect of Ribavirine on vaccinia virus :

In order to make a clinical trial with a recombinant vaccinia virus, we have to test the ability of an antiviral compound to inhibit the development of vaccinia lesions. As we have previously seen, when mice are injected with VV5021X5065, they do not develop vaccinia lesions (even immunodeficient mice). Therefore, the antiviral efficiency was tested in nude mice injected with VVwt and treated with Ribavirine. The table 3 shows the number of mice with lesions after antiviral treatment, and shows that an *in vivo* efficient anti-vaccinia compound is available.

3- *In vivo* efficacy of VV5021X5065:

The viral construct VV5021X5065 was tested for *in vivo* anti-tumoral activity against cells expressing the E7 molecule, and was compared to VV5021X5061 (E6*E7*L1L2), VVIL2 and VVcontrol (186). This activity was assessed in 2 models : immunoprotection and immunotherapeutic experiments, whose protocols are similar in all experiments described in this report.

3.1 - Immunoprotection experiment : Mice were immunized subcutaneously 3 times with 10^7 pfu of VV5021X5065. Three days after the last immunization, they were challenged with 10^3 E7W1 cells. The percentage of surviving animals in function of time is represented in figure 1 and shows a clear increase for mice immunized with VV5021X5065.

	10^6 pfu	10^7 pfu	10^8 pfu
VVwt	0/5*	1/5	1/5
VV5021X5065	0/5	0/5	0/5

Table 1 : Toxicity of VV5021X5065 compared to VVwt - Dose effect. *number of mice with lesions .

	IV	IP	IM	IC
VVwt	5/5 (2 dead D16)	3/5	1/5	4/5 (4 dead D5)
VV5021X5065	0/5	0/5	0/5	0/5

Table 2 : Toxicity of VV5021X5065 compared to VVwt - Immunization route effect. *number of mice with lesions.

	control		Ribavirine (30mg/kg)		Ribavirine (300mg/kg)	
n° mice	fingers	tail	fingers	tail	fingers	tail
1	0	0	++++	+	0	0
2	0	+	0	+	0	0
3	+++	+++	+++	+	0	0
4	+++	+++	0	0	0	0
5	0	+	+	+	0	0
6	++	+++	+++	++	0	0
7	0	+	++++	++	0	0
8	0	0	+++	++	0	0
9	+++	+++	+	0	0	0
10	+++	+++	0	0	0	0

Table 3 : *In vivo* antiviral effect of Ribavirine on wild type vaccinia virus lesion in nude mice.

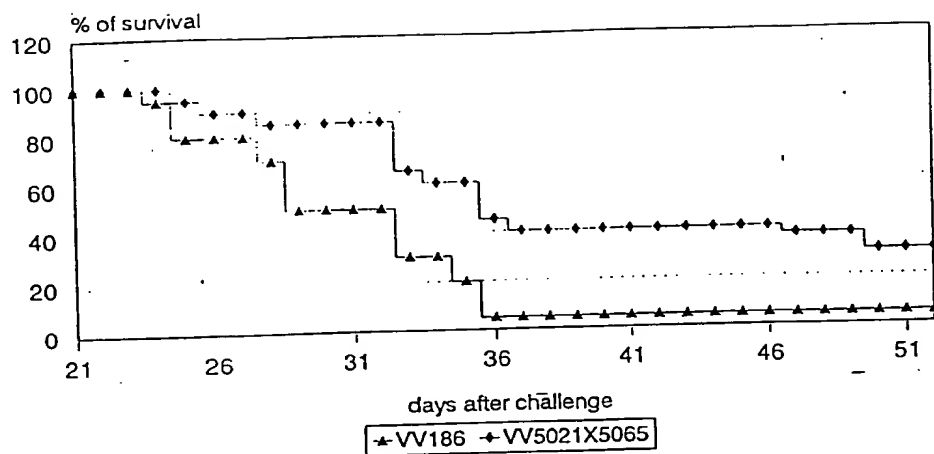
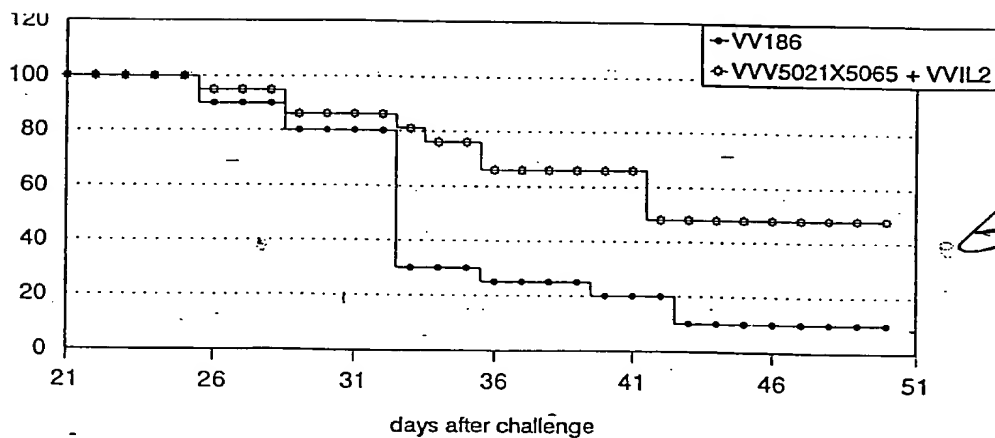
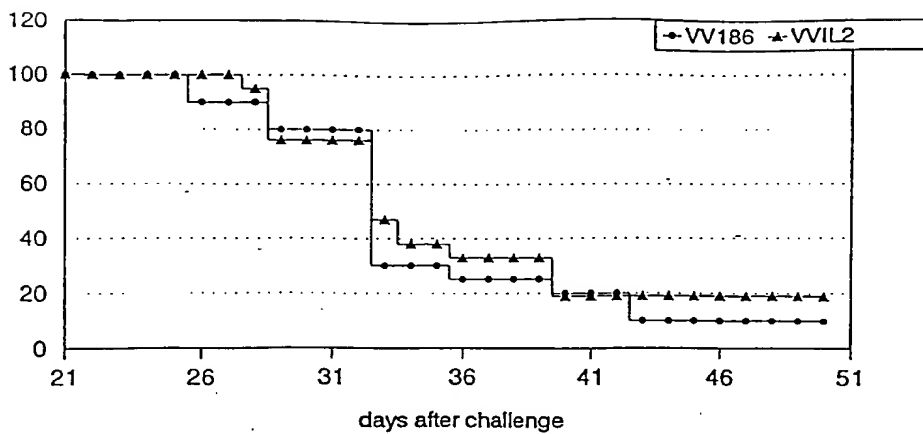


Figure 1 : In vivo efficacy of VV5021X5065. Immunoprotection experiment. 10^7 pfu VV SC, 3X, 10^3 E7W1 SC.



← + combinaison de virus

Figure 7 : Immunotherapy experiment. Effect of IL2 dosage. Mice were injected with 10^3 E7W1 cells and then treated 3 times with 10^7 pfu of VV (SC).

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